

Introduction dataset – Day 1

Invasive disease caused by *Streptococcus pneumoniae* and patient mortality

Aldert Zomer – Bas Dutilh



Streptococcus pneumoniae



Sinusitis
mastoiditis
Bronchitis

Meningitis

Otitis media

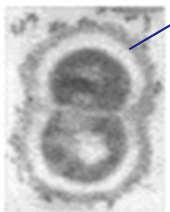
Capsule

Vaccin
PCV7,10,13
2006

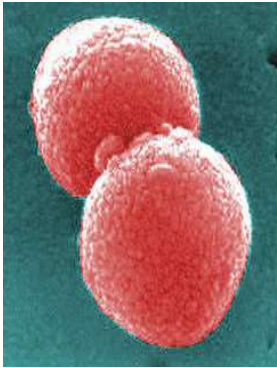
Septic arthritis

Bacteremia
Sepsis

Pneumonia



Cohort



S.pneumoniae



Blood culture +



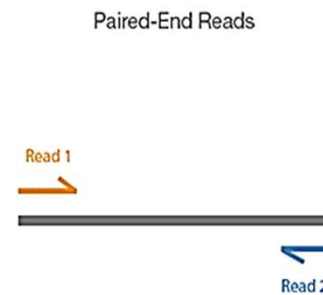
Two Dutch hospitals `2001-2011`



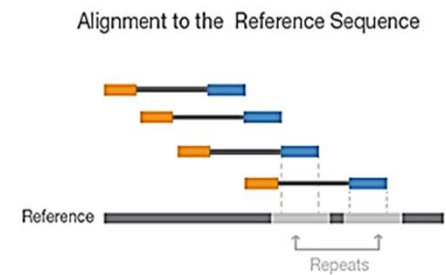
Clinical data



Serotype



Illumina PE sequencing: Sanger Institute

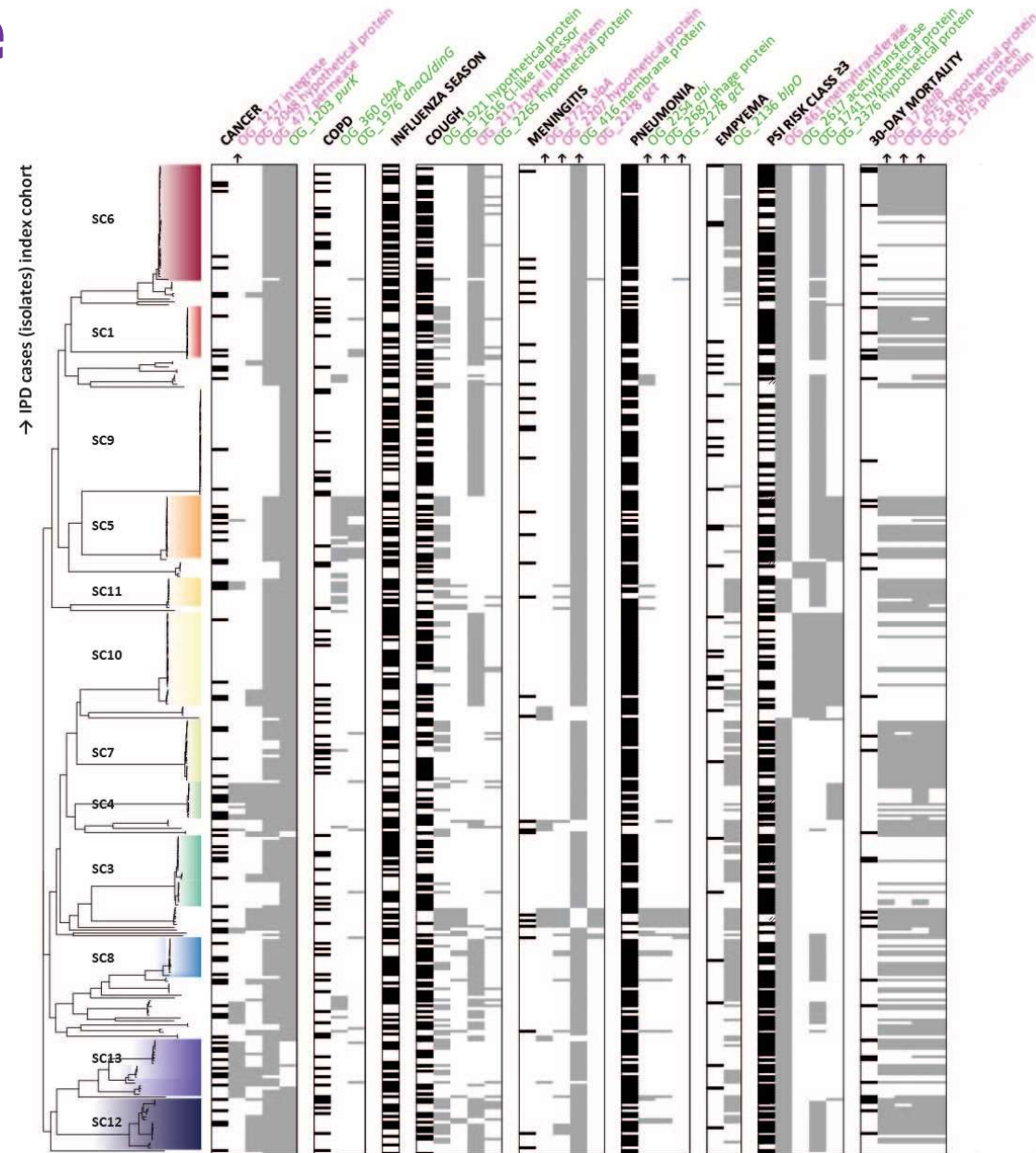


Severity of disease

→ Presence of phenotypes and associated orthologous genes

Are there specific bacterial clones that are associated with patient mortality?

What pneumococcal virulence genes are over-represented in the isolates from patients that died within 30 days of hospital admission and could be associated with patient mortality



CLINICAL IPD PHENOTYPE
 Positively associated OG
 Negatively associated OG
 ↑ Selected for validation





Universiteit Utrecht

Bacterial GWAS

Linking phenotype to genotype in bacteria

Aldert Zomer

Microbial Genomics

2020 – Day 2

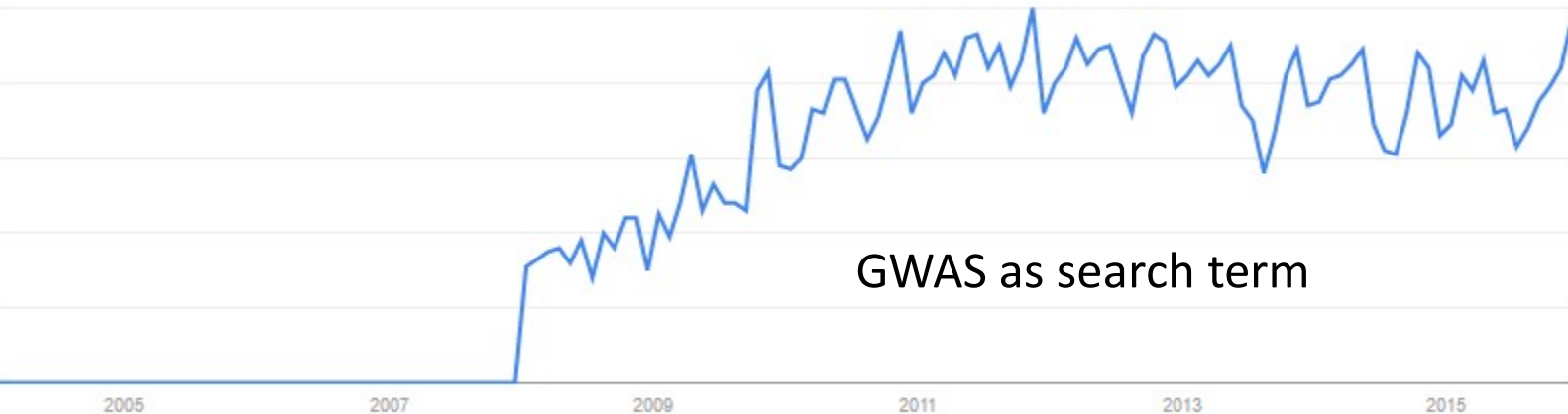
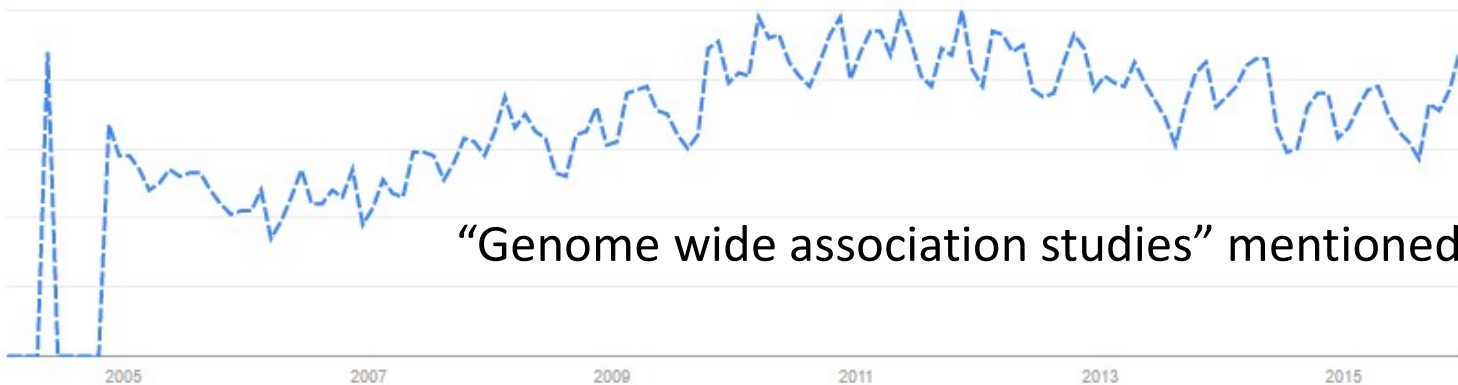
GWAS

- **G**enome **W**ide **A**ssociation **S**tudies:
- Linking a phenotype to a genotype
- Phenotype: Trait, disease
- Genotype: (combinations of) Single Nucleotide Polymorphisms (SNPs), gene variants, complete genes



History

- HapMap Project (2002, 2005, 2007, 2009)
- The 1000 Genomes Project (2008)



Google Trends

Basic idea

Genotype individuals/species/isolates for a large number of SNPs spread in a generally unspecified way throughout the genome. Look for association.

SNPs →

patients ↓

2	1	0	1	2	1	1	0	0	0	2	0	Control
0	1	1	0	1	2	0	1	0	0	2	1	Control
0	0	0	2	0	0	0	0	0	2	1	0	Control
0	1	1	2	1	0	1	1	1	1	2	2	Control
2	0	2	1	0	1	1	0	0	0	2	2	Control
1	1	2	1	2	2	0	1	0	0	1	1	Control
1	1	0	2	1	1	0	0	1	0	0	1	Control
0	0	1	0	2	1	0	1	2	0	1	1	Case
0	2	2	0	0	1	1	1	2	1	0	0	Case
0	0	0	2	0	2	2	0	2	2	1	2	Case
0	1	1	0	0	0	1	1	2	2	1	0	Case
2	0	2	1	1	2	2	0	2	0	2	2	Case
1	2	0	1	2	0	0	0	2	1	1	2	Case
1	1	0	0	2	2	2	0	2	0	2	0	Case

What do you see in the table? (hint: diploid)



Basic idea

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SNPs →

patients ↓

2	1	0	1	2	1	1	0	0	0	2	0	Control
0	1	1	0	1	2	0	1	0	0	2	1	Control
0	0	0	2	0	0	0	0	0	2	1	0	Control
0	1	1	2	1	0	1	1	1	1	2	2	Control
2	0	2	1	0	1	1	0	0	0	2	2	Control
1	1	2	1	2	2	0	1	0	0	1	1	Control
1	1	0	2	1	1	0	0	1	0	0	1	Control
0	0	1	0	2	1	0	1	2	0	1	1	Case
0	2	2	0	0	1	1	1	2	1	0	0	Case
0	0	0	2	0	2	2	0	2	2	1	2	Case
0	1	1	0	0	0	1	1	2	2	1	0	Case
2	0	2	1	1	2	2	0	2	0	2	2	Case
1	2	0	1	2	0	0	0	2	1	1	2	Case
1	1	0	0	2	2	2	0	2	0	2	0	Case

homozygous for mutation: associated with case



Basic idea (2)

	SNP present	SNP absent
with phenotype	20	3
without phenotype	4	16

$$p = 1.19 \times 10^{-5}$$

2x2 (or 3x2 in diploid genomes) contingency tests

e.g.

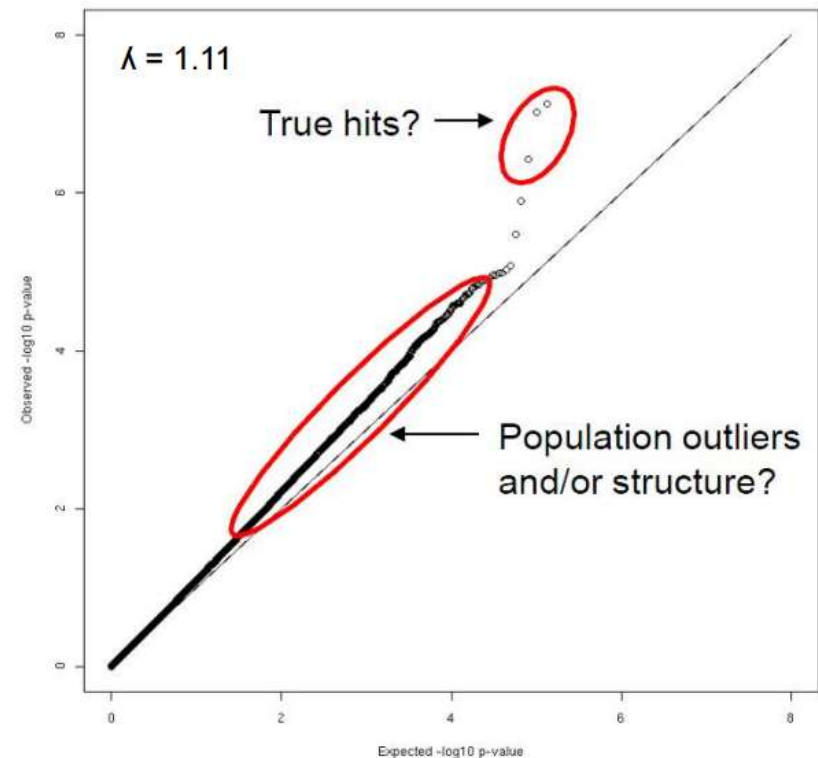
Fisher exact (small samplesizes, values <10)

Chi squared (large samplesizes, values >10)

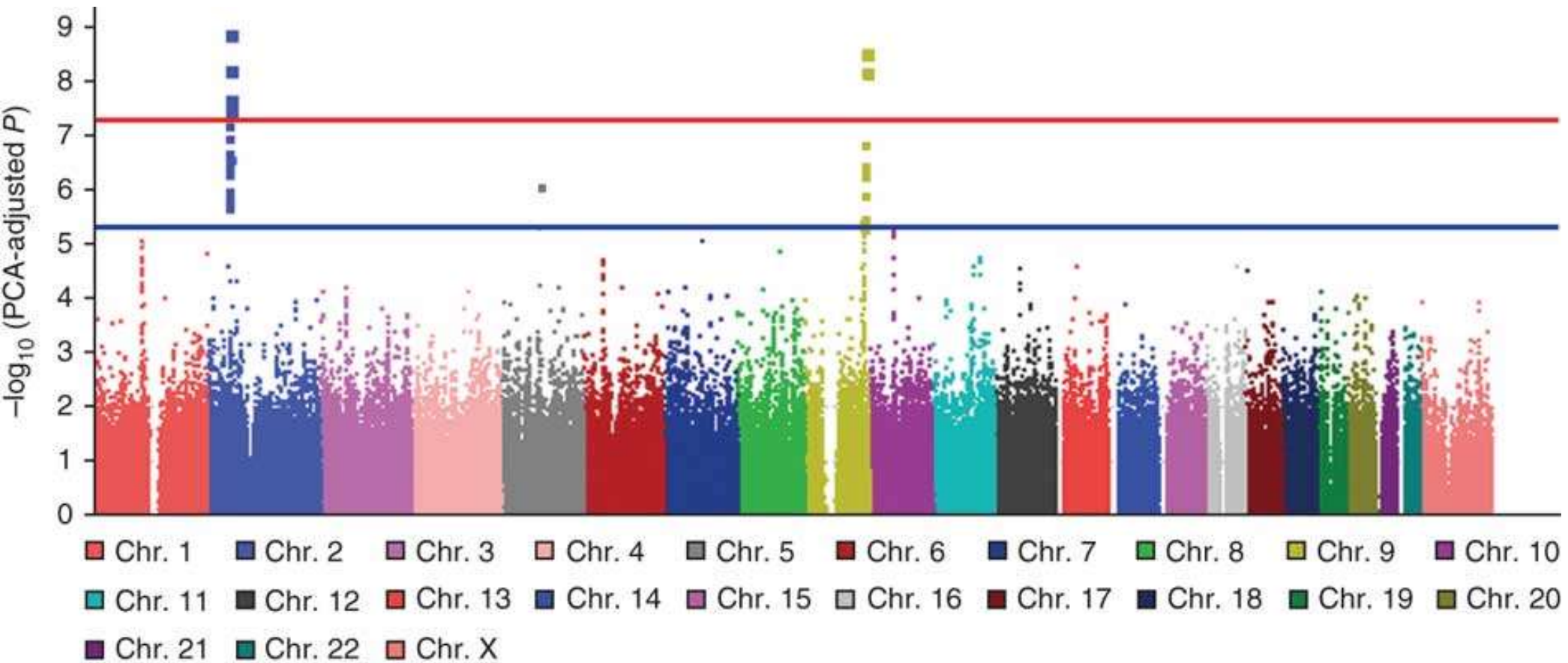
QQ-plot:

Plot the expected p-values against the observed p-values

Strong deviations are likely candidates



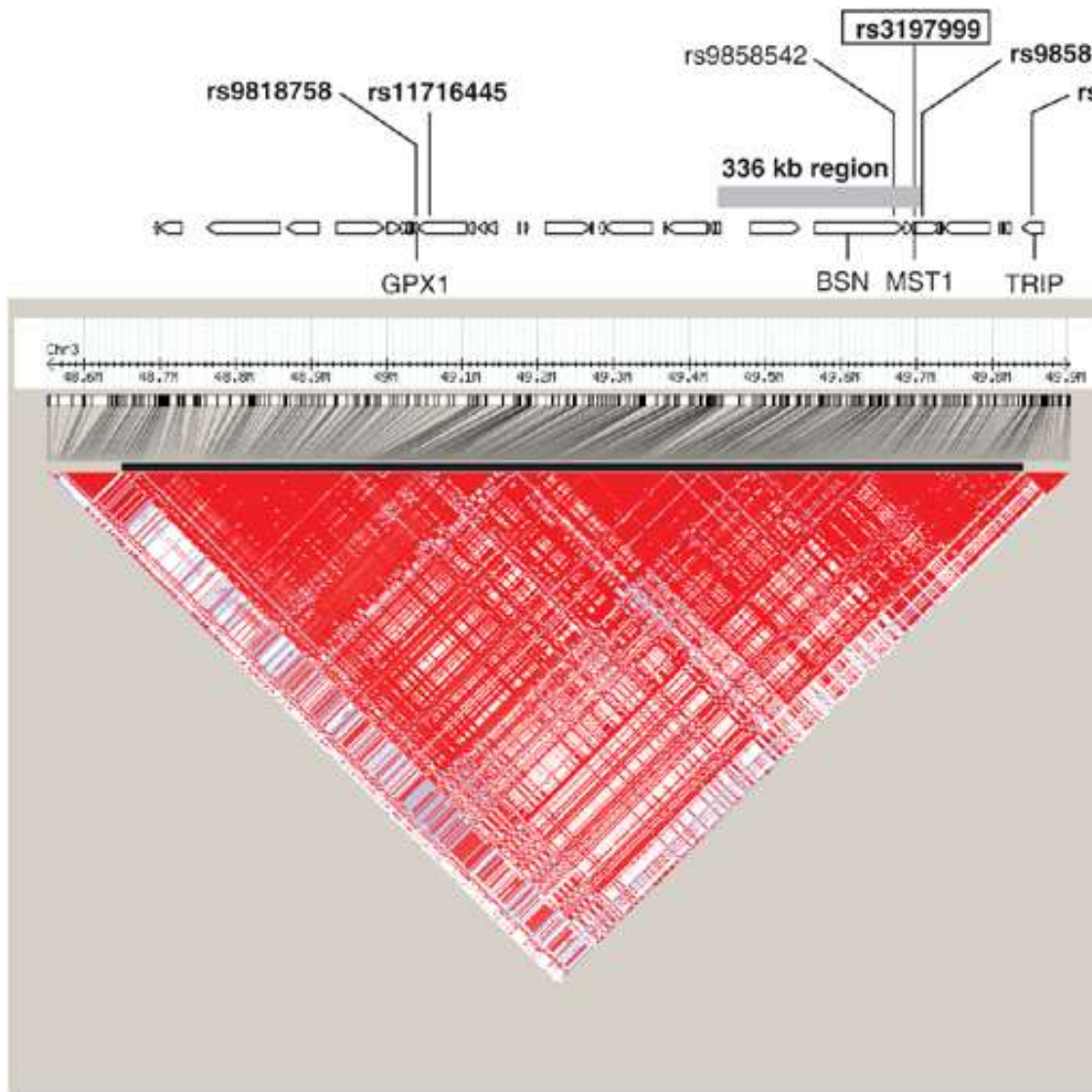
Basic idea (3)



Negative log₁₀ P-values plotted against location on genome: Manhattan plot



Basic idea (4)



Linkage disequilibrium plots are used to visualize alleles that co-occur more often than what would be expected if alleles were independently, randomly sampled, based on their individual allele frequencies.



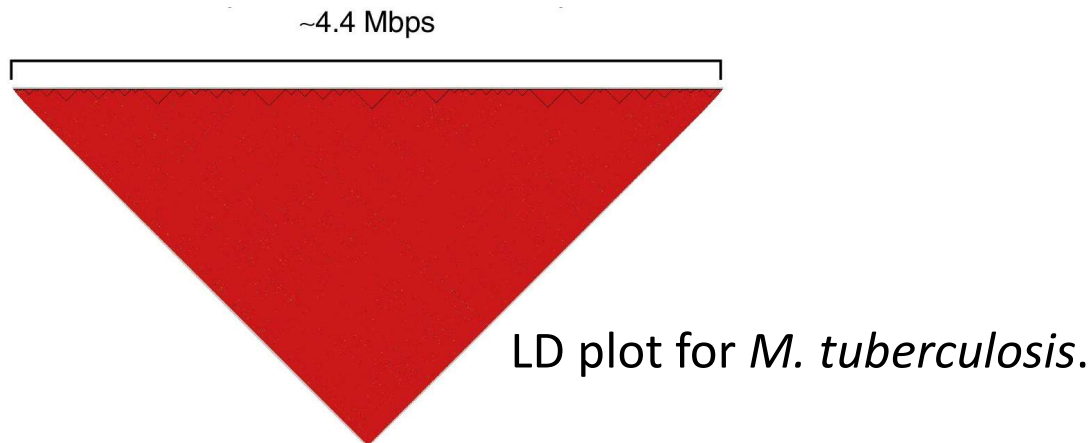
Population structure correction

Population structure: Potentially a problem in human genetics.
A real problem in bacterial genetics

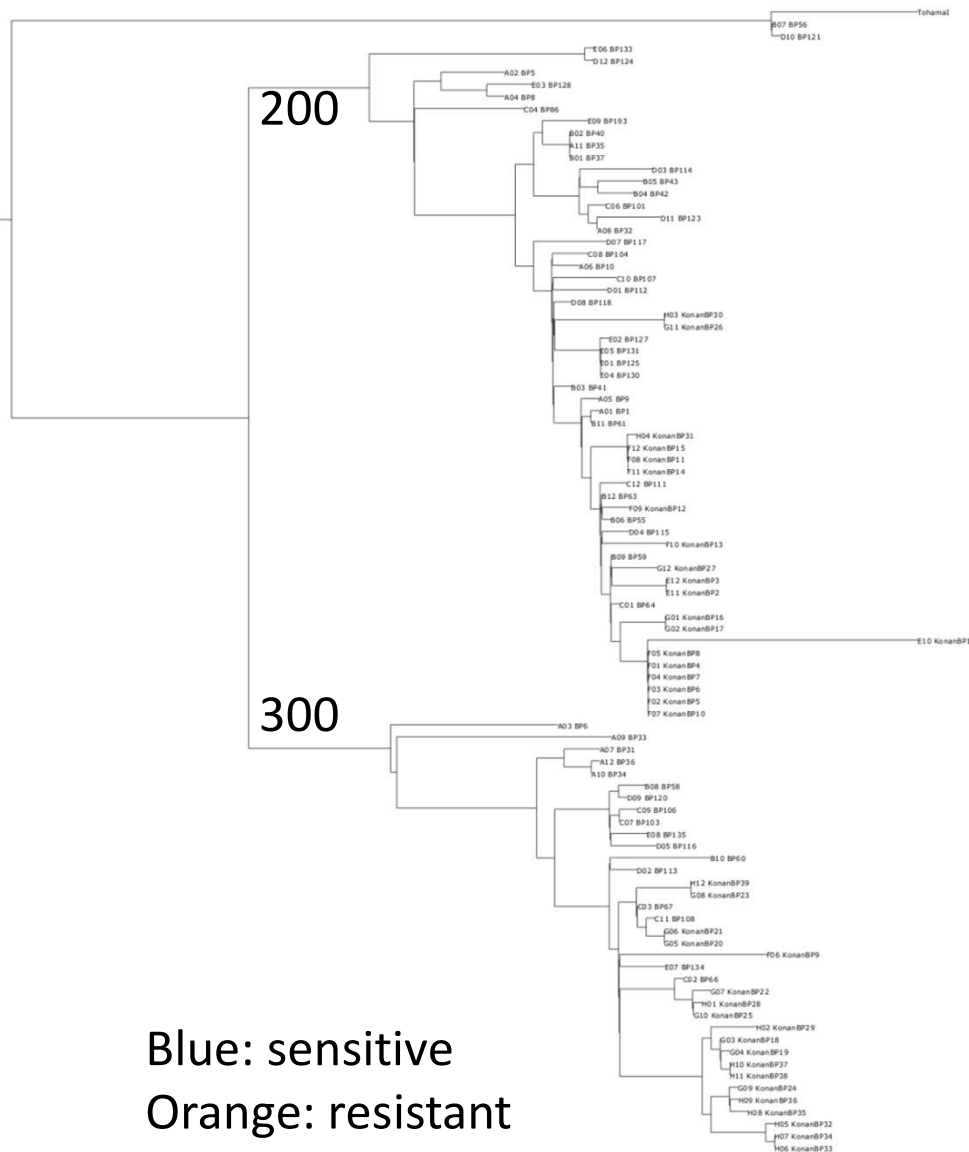


Population structure correction

- Population structure (in humans) occurs through mechanisms such as genetic drift, ancestral divergence and non-random mating
- Confounds GWAS: higher than expected allele frequencies within certain members of the study set
- Problematic in bacterial GWAS: haploid and asexual. In the absence of recombination, all fixed genetic variants will be passed on to descendants and be in linkage disequilibrium with other mutations that occur in that lineage.



Population structure correction



Example:

Find the SNP associated with antimicrobial resistance

But.. Resistance against an antibiotic is primarily associated with a certain branch in the phylogenetic tree.

Standard contingency test will associate phylogenetic markers with resistance, 100s of SNPs (clade 3 defining SNPs)



Population structure correction

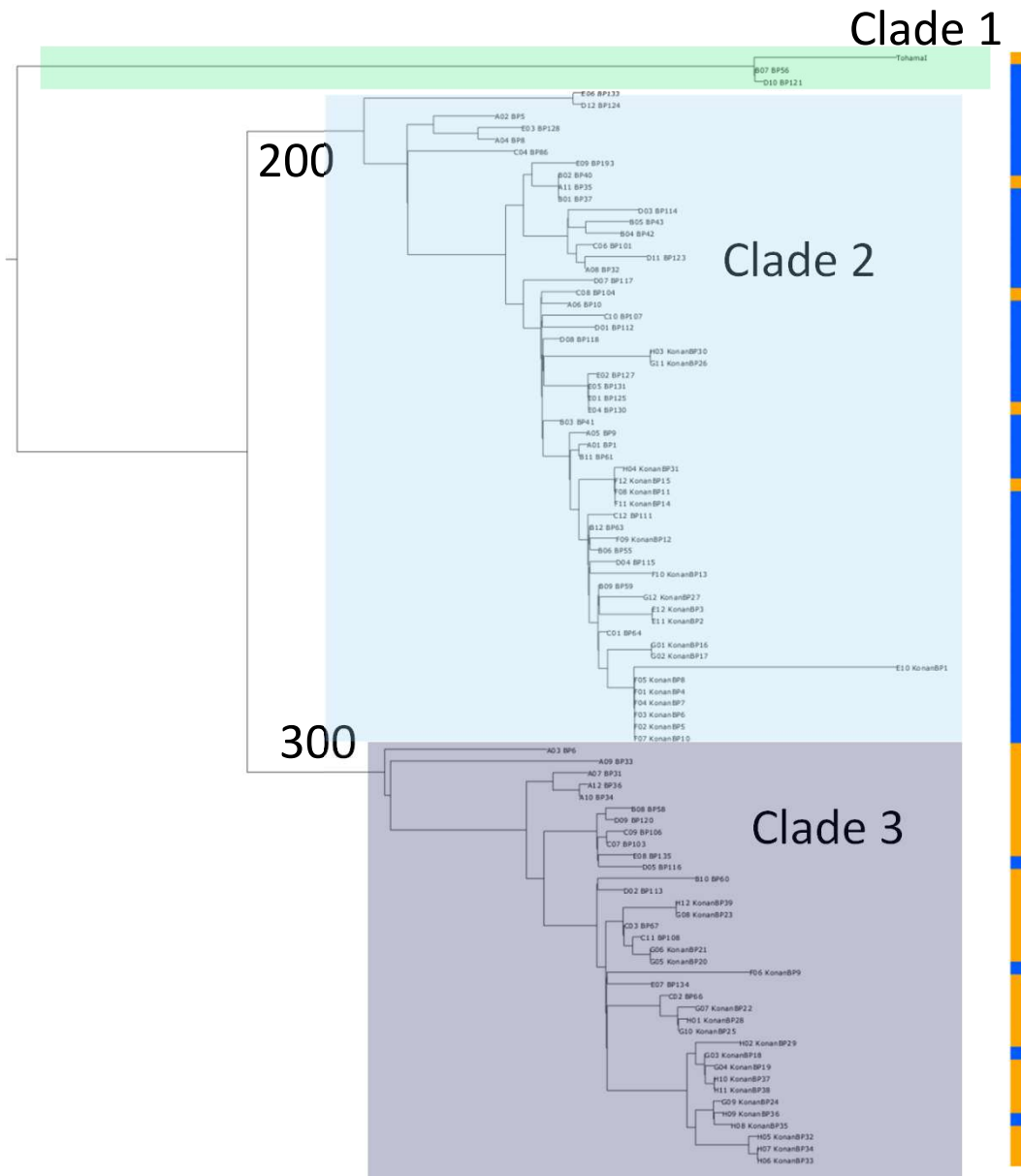
Determine population groups:

- Pre-existing knowledge from e.g. MLST
- multi-dimensional scaling in PLINK
- principal component analysis in EIGENSTRAT
- Bayesian analysis of genetic population structure: BAPS
- Infer clones based on branch lengths in phylogenetic tree
- Many others..

Use the groups as covariates in association testing (e.g. with the Cochran-Mantel-Haenszel test)



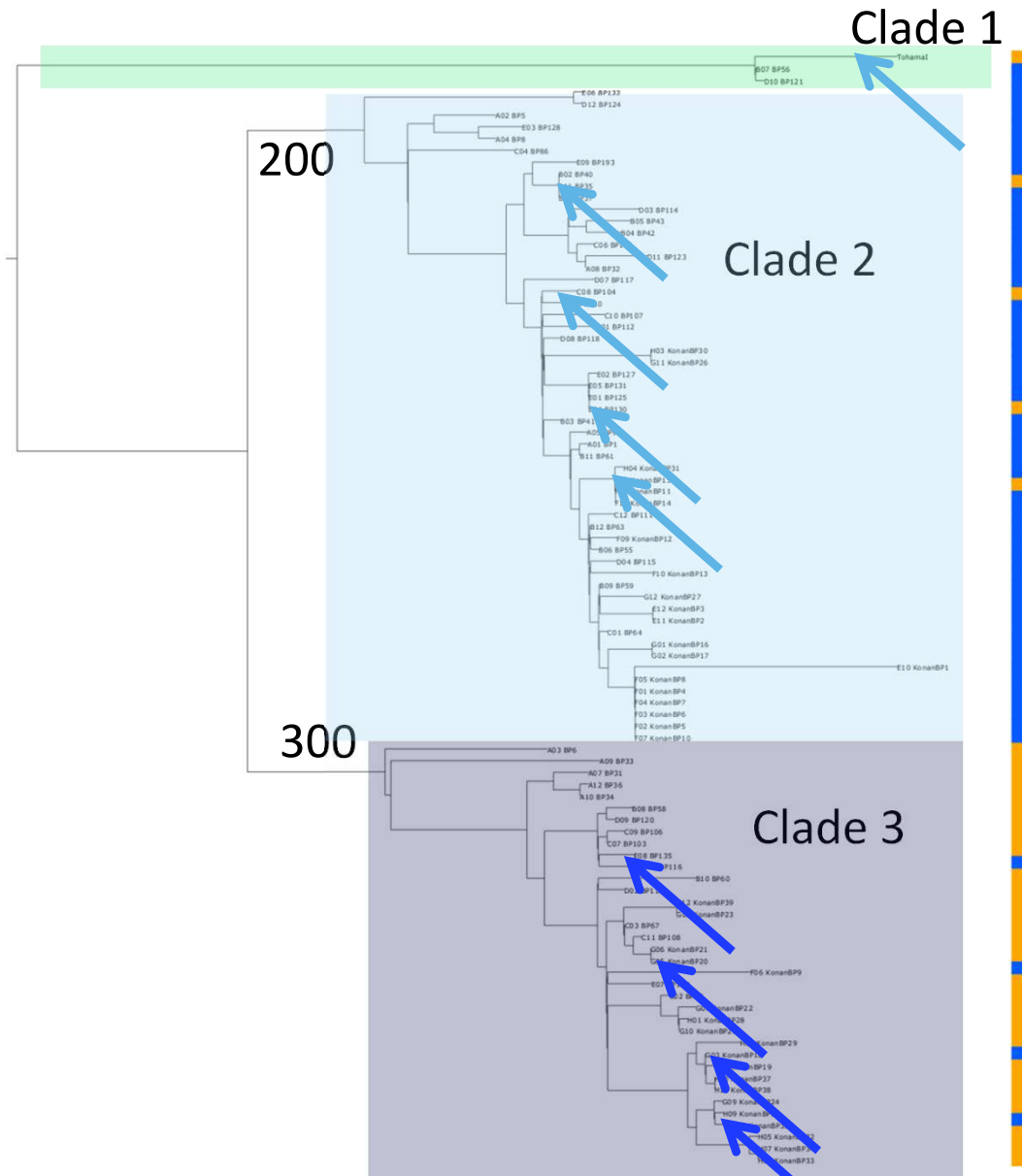
Population structure correction



Cochran-Mantel-Haenszel:

- Performs association testing per clade
- Computes a weighted p value

Population structure correction



Cochran-Mantel-Haenszel:

- Performs association testing per clade
- Computes a weighted p value

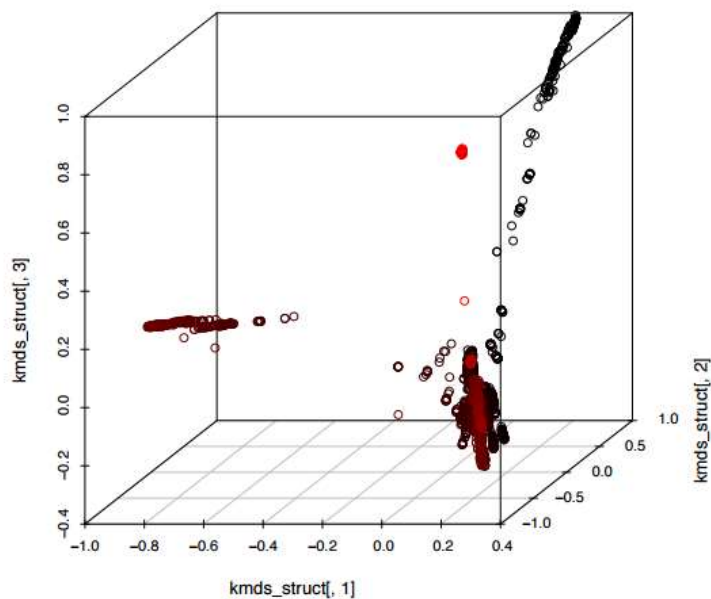
Alternatively:

Count repeated and independently emerged mutations occurring more often on branches of cases relative to controls

(PhyC: Farhat et al Nat Genet. 2013)

Implemented in Scoary

Newer methods



MENU ▾

nature
COMMUNICATIONS

Altmetric: 18 Views: 1,806

[More detail >>](#)

Article | [OPEN](#)

Sequence element enrichment analysis to determine the genetic basis of bacterial phenotypes

John A. Lees, Minna Vehkala, Niko Vähimäki, Simon R. Harris, Claire Chewapreecha, Nicholas J. Croucher, Pekka Marttinen, Mark R. Davies, Andrew C. Steer, Steven Y. C. Tong, Antti Honkela, Julian Parkhill, Stephen D. Bentley & Jukka Corander [✉](#)

SEER: Also infers population structure automatically and uses position in an nMDS plot as cofactor. Nat Commun. 2016 Sep 16;7:12797

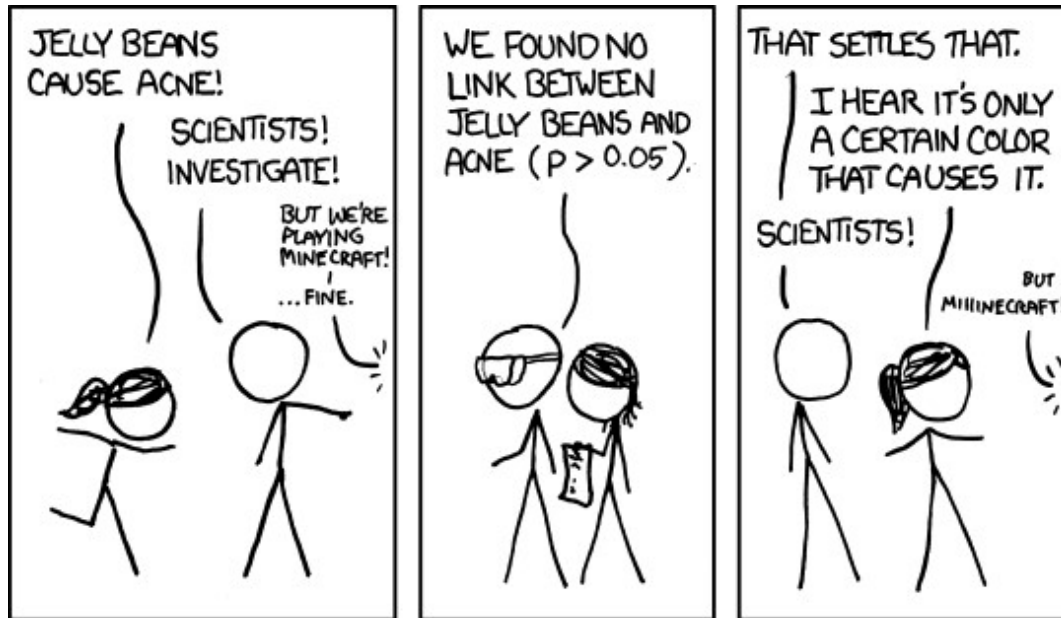


Multiple testing correction

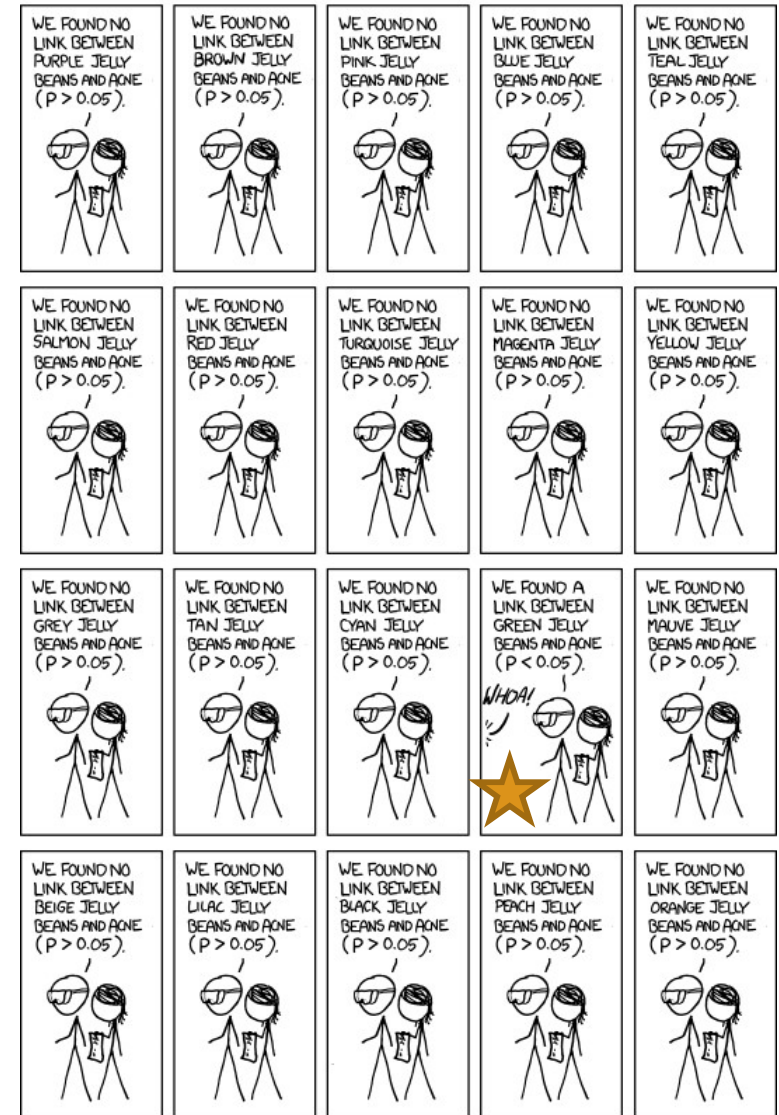
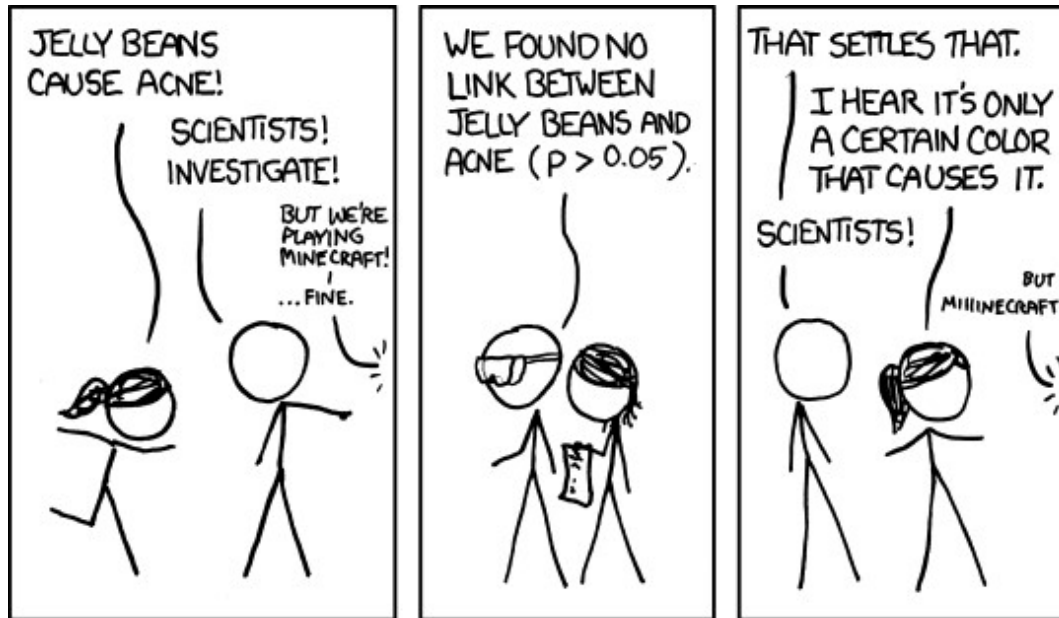
1000 SNPs have a p-value < 0.05 . Are they all true positives?



Multiple testing problem



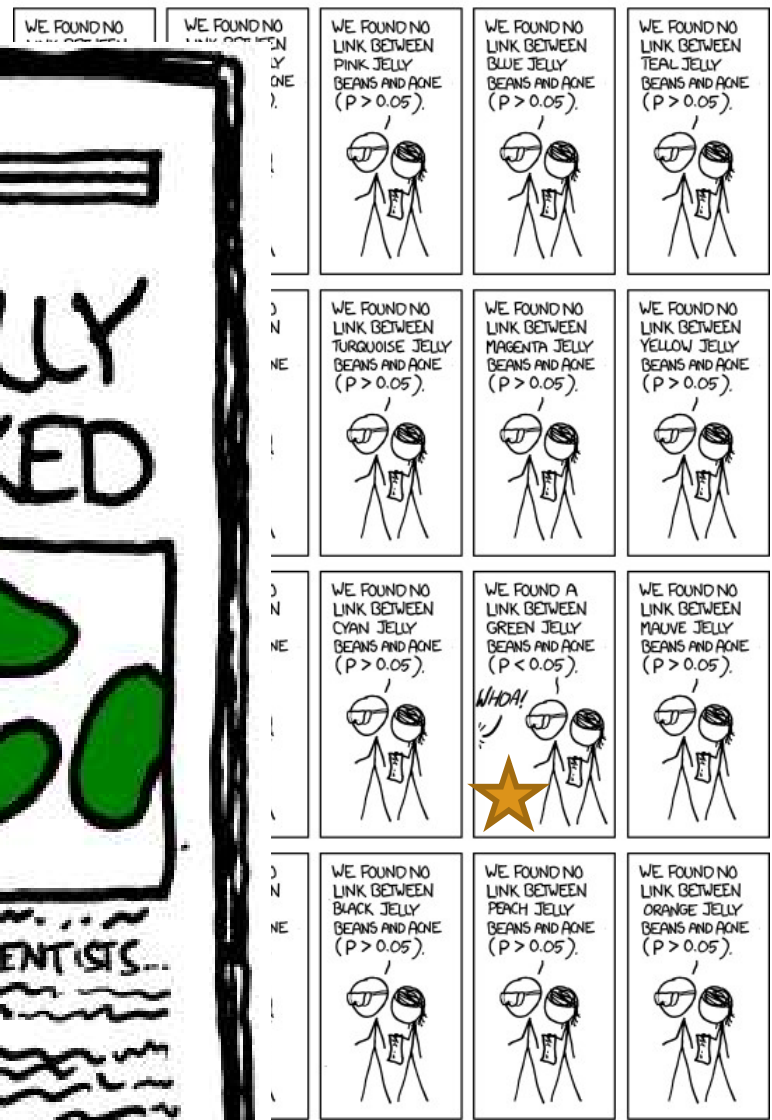
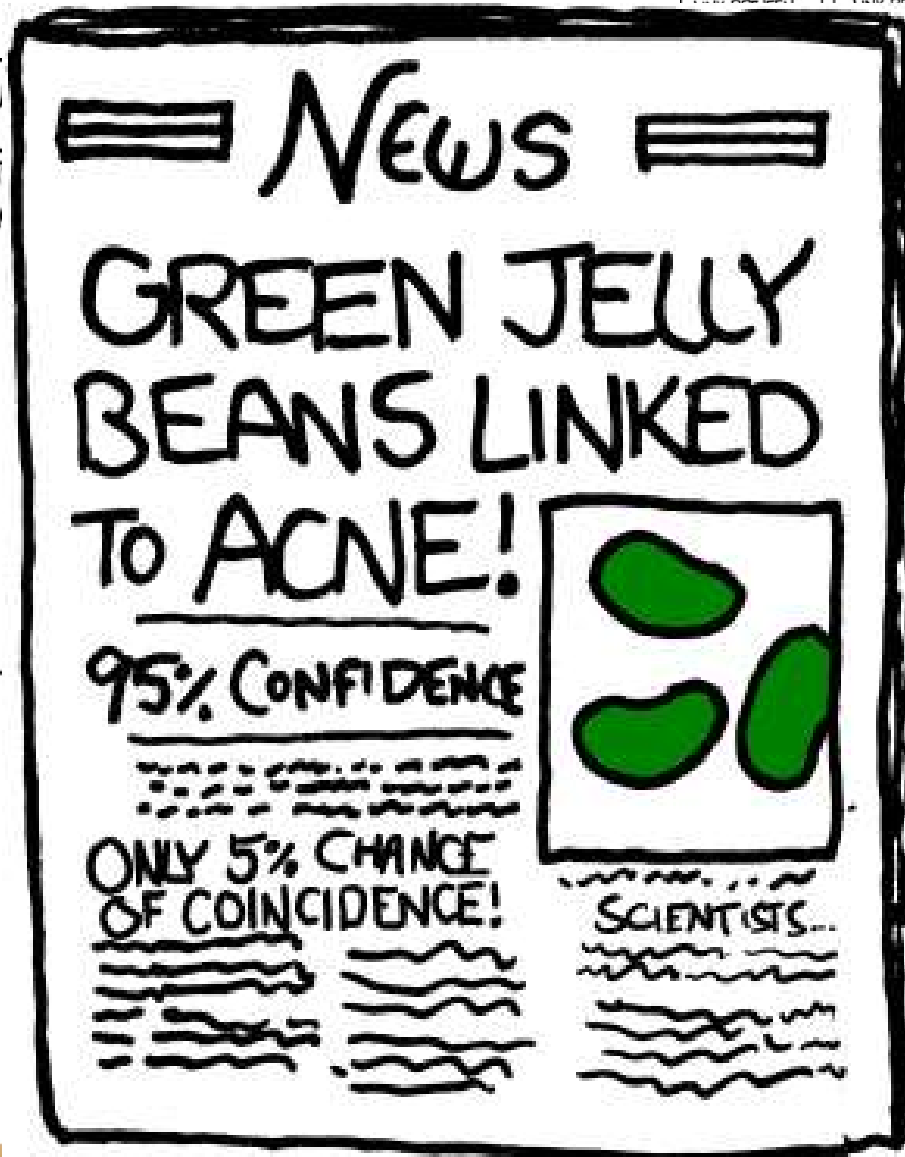
Multiple testing problem



Multiple testing problem



W
L
J
A



Multiple testing: Adjusting

- Significance threshold must adjust for Type I error (a false positive); spurious statistical significance arising from multiple comparisons involving hundreds of thousands of SNPs

Dudbridge F, Gusnanto A (2008) Estimation of significance thresholds for genome-wide association scans. *Genetic Epidemiology* 32:227-34

Pe'er I, Yelensky R, Altshuler D, Daly MJ, (2008) Estimation of the multiple testing burden for genome-wide association studies of nearly all common variants. *Genetic Epidemiology*, May;32(4):381-5



Multiple testing: Adjusting

- Bonferroni correction
- Benjamini Hochberg (false discovery rate)
- Permutation – computationally demanding
- Bayesian approaches - computationally demanding

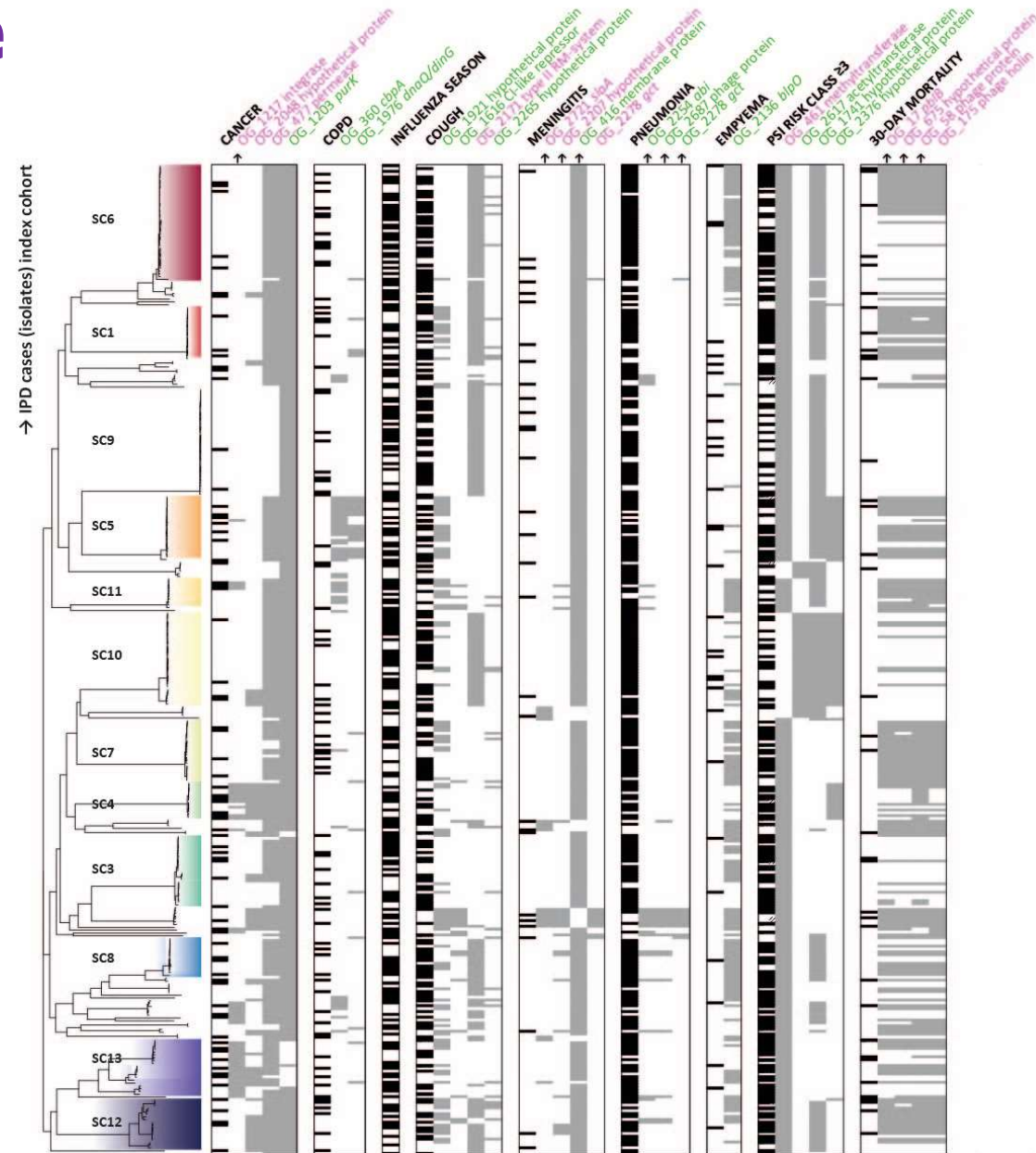


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